

Immobilized PNGase F



Removal of N-glycans from Glycoproteins

SmartEnzymes™





Hydrolysis of N-glycans



Immobilized PNGase F is a resin with the PNGase F enzyme covalently coupled to agarose beads for removal of N-glycans on antibodies, fusion proteins and other N-glycosylated proteins. The enzyme is widely used for sample preparation prior to MS analysis – to reduce the protein heterogeneity and enable released glycan analysis – and to study the functional role of the N-glycan.



N-linked glycans on glycoproteins





15+ min denaturing or 1+h native



Deglycosylation Workflow



PNGase F (Peptide N-glycosidase F) is a glycoamidase hydrolyzing the amide bond between the polypeptide asparagine and the innermost GlcNAc of all mammalian asparagine-linked complex, hybrid, or high-mannose oligosaccharides.

Immobilized PNGase F

During the reaction, the asparagine residue from which the glycan is removed is deamidated to aspartic acid. The glycoprotein sample is incubated with the Immobilized PNGase F resin in a spin column for 1 h to overnight using non-denaturing conditions, or 15-60 min using denaturing conditions and the deglycosylated glycoprotein is then easily collected by a centrifugation step.

Product Formats



Immobilized PNGase F Hydrolysis of N-glycans from glycoproteins in spin columns under native conditions



Immobilized PNGase F Denaturing Hydrolysis of N-glycans from glycoproteins in spin columns under denaturing conditions

Immobilized PNGase F		Ų
PRODUCT	DESCRIPTION	ID
Immobilized PNGase F	5×0.2 mg microspin columns	G1-PF6-010
Immobilized PNGaseF	10 × 0.2 mg microspin columns	G1-PF6-020

Removal of N-glycans under Native Conditions using Immobilized PNGase F

Removing the Fc N-glycan with PNGase F under native conditions enables characterization of the free N-glycan as well as the function and structure of the deglycosylated antibody. Trastuzumab was used to demonstrate the efficient removal of Nglycans by Immobilized PNGase F under native reaction conditions. The mass shift demonstrates successful removal of the Fc N-glycans with no enzyme interfering in the analysis as compared to the sample processed with PNGase F in solution.



Removal of N-glycans under native conditions. TIC chromatogram (left) and deconvoluted mass spectra (right) of the Fc/2 fragment of trastuzumab treated with Immobilized PNGaseF or PNGaseF in solution.

Rapid N-glycan Removal using Immobilized PNGase F Denaturing



The deglycosylation ability of Immobilized PNGase F on a selection of glycoproteins is here demonstrated. For example, the commonly used glycoprotein standard RNaseB was completely deglycosylated in 15 min using Immobilized PNGaseF Denaturing.

	ABATACEPT	RNASE B	CETUXIMAB
Protein type	Fusion protein	Glycoprotein	lgG1
N-glycans	6	1	4
O-glycans	8	-	-
Reaction conditions	Native, 18h or denaturing, 30 min	Denaturing and reducing, 15 min	Native, 1 h or denaturing, 15 min
Preparation of samples for analysis	Reduced	N/A	FabRICATOR digested





Key characteristics and deconvoluted mass spectra or SDS-PAGE assay of a selection of glycoproteins deglycosylated using Immobilized PNGase F. Abatacept was analyzed by reverse-phase LC-MS and RNase B and cetuximab were analyzed on SDS-PAGE. The mass shifts in the deconvoluted mass spectra or on the gels show the successful removal of N-glycans from the various glycoprotein substrates.

Immobilized PNGase F Denaturing		
PRODUCT	DESCRIPTION	ID
Immobilized PNGase F Denaturing	5×0.2 mg microspin columns + 5×1 mg <i>Rapi</i> Gest [™] SF	G2-PDK-010
Immobilized PNGase F Denaturing	10 × 0.2 mg microspin columns + 10 × 1 mg <i>Rapi</i> Gest [™] SF	G2-PDK-020

RapiGest[™] SF Surfactant included in Immobilized PNGase F Denaturing. RapiGest[™] SF Surfactant is a trademark of Waters Corporation.

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